### **Model Selection**

Michael Noonan

Biol 520C: Statistical modelling for biological data



- 1. The Problem of Overfitting
- 2. Likelihood-Ratio Tests
- 3. Information Criteria
- 4. IC Considerations

#### The Problem of Overfitting



We started by fitting simple linear regression models of the form:

$$y_i = \beta_0 + \beta x_i + \varepsilon_i$$

We then extended this to multiple linear regression of the form:

$$y_i = \beta_0 + \beta_1 x_{1i} + \beta_2 x_{2i} + \ldots + \beta_n x_{ni} + \varepsilon_i$$

Last lecture we covered linear mixed effects model that add additional structure to account for correlations within groups:

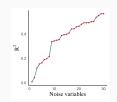
$$\mathbf{y}_{i} = X_{i}\beta + Z_{i}\mathbf{b}_{i} + \varepsilon_{i}$$

#### The Problem of Overfitting

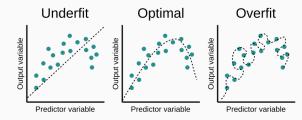
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Lecture by lecture we've been increasing the complexity of our models

But more complexity does not necessarily mean improved performance, or that the parameters are meaningful.



This puts us in a situation where we need to strike an optimal balance between having too many or too few features in our models.





Walters & Ludwig (1981) simulated fish population dynamics using complex age-structured model (different births and deaths for fish of different ages).



When data were realistically sparse and noisy a simple model without age structure resulted in the best predictions, even though they knew for a fact that there were age differences (!)

**Why?** Data contain a fixed amount of information (limited by resources, time, instrument precision, etc.) and as we estimate more and more parameters that information gets spread thinner and thinner.

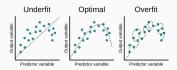
For Walters & Ludwig, spreading the information across age classes meant each age class' dynamics were included, but poorly estimated.



An **underfit** model fails to accurately predict the data that were used to fit the model, and test datasets or future conditions.

An **overfit** model gives a very low prediction error on the dataset used to fit the model, but has a very high prediction error on test data.

This happens because you're fitting the noise not the signal.







In practice, adding features/complexity to a model usually boils down to answering the question:

Does adding an extra parameter improve the fit sufficiently to justify the additional complexity?

... or seen another way:

Is there enough information in my data to support the additional complexity that comes with an extra parameter?

Note: Excluding a parameter does not necessarily mean an effect does not exist. It does mean we can't estimate it in a meaningful way from the information we have on hand.



The goal of **modelling** is to identify the simplest model possible that captures all of the most important features of our data/system.

The goal of **model selection** is to know whether adding another parameter to our model not only improves the performance, but improves it by some specific amount (i.e., not just marginal gains) in order to minimise the risk of overfitting. If the more complex model doesn't pass this threshold, then it is rejected in favour of the simpler model.

Model selection highly controversial, but extremely important. We're going to cover two approaches:

- 1. Likelihood-ratio tests
- 2. Information criteria

### Likelihood-Ratio Tests



The likelihood-ratio test compares a pair of **nested** models based on the ratio of their likelihoods.

$$\lambda_{\mathsf{LR}} = -2 \ln \left[ \frac{\mathcal{L}(\text{Reduced model})}{\mathcal{L}(\text{Full model})} \right]$$

The likelihood-ratio test statistic is often expressed as a difference between the log-likelihoods

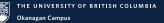
$$\lambda_{\mathsf{LR}} = -2(\mathsf{ln}[\mathcal{L}(\mathsf{Reduced})] - \mathsf{ln}[\mathcal{L}(\mathsf{Full})])$$



A 'simple' model is nested in a 'complex' model if the full model reduces to the simpler model when parameters are set to some fixed values (usually 0, 1, or  $\infty$ ).

For example:

 $\begin{array}{ll} y_i = \beta_0 + \beta_1 x_{1i} + \beta_2 x_{2i} + \varepsilon_i & y = \frac{ax}{1 + (\frac{b}{b}x)} \mbox{ (Beverton-Holt)} \\ \mbox{if we set } \beta_2 \mbox{ to } 0, \mbox{ then:} & \mbox{if we set } b \to \infty, \mbox{ then:} \\ y_i = \beta_0 + \beta_1 x_{1i} + \varepsilon_i & y = ax \mbox{ (Linear model)} \\ \mbox{So the models are nested} & \mbox{ Again the models are nested} \end{array}$ 



A 'simple' model is nested in a 'complex' model if the full model reduces to the simpler model when parameters are set to some fixed values (usually 0, 1, or  $\infty$ ).

For example:

 $y_i = \beta_0 + \beta_1 x_{1i} + \varepsilon_i$  is not nested in  $y_i = \beta_0 + \beta_2 x_{2i} + \varepsilon_i$ 

because if we set  $\beta_1$  to 0, then:

 $y_i = \beta_0 + \varepsilon_i$ 

Both are simple linear models, and might even be based on the same data, but they are not nested.



Both the deterministic and stochastic components of models can be nested.

For example:

The Poisson distribution is nested in the negative binomial distribution. The binomial distribution is nested in the beta-binomial distribution

In other words, likelihood ratio tests can be used to identify both deterministic *and* stochastic components of your model (we'll cover non-gaussian models later, but keep this in mind).



# So how does being able to quantify $\lambda_{\rm LR}$ help us identify the best model structure?

According to Wilks' theorem, as the sample size *n* approaches  $\infty$ , the test statistic  $\lambda_{LR}$  will be chi-squared distributed with degrees of freedom equal to difference in the number of parameters between the two models.

This implies that we can compare  $\lambda_{\rm LR}$  to the  $\chi^2$  value corresponding to a desired statistical significance threshold (usually  $\alpha = 0.05$ ) as an approximate statistical test.



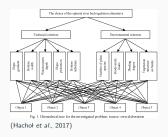
```
library(nlme)
data <- read.csv("Ant Richness.csv")</pre>
FIT <- gls(num_sp ~ latitude + elevation, data = data, method = "ML")
FIT Reduced <- gls(num sp ~ latitude, data = data, method = "ML")
lambda <- -2*(FIT_Reduced$logLik - FIT$logLik)
pchisg(lambda, df = 1, lower.tail=FALSE)
0.001948725
anova(FIT, FIT Reduced)
            Model df
                          ATC
                                   BIC
                                       logLik Test L.Ratio p-value
FIT
                1 4 16.11769 20.48186 -4.058844
FIT Reduced
                2 3 23,71490 26,98803 -8,857452 1 vs 2 9,597217 0,0019
```

This is telling us that the extra parameter is resulting in a significant improvement to the model, so the extra complexity is worth the cost.

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With only a small number of nested models to compare, likelihood ratio tests are quick easy, and statistically efficient.

As the complexity of the full model increases, the number of pairwise LRTs increases dramatically and soon becomes unwieldy.



If some of the models aren't nested in one another, there is no way to compare them using LRTs.

## **Information Criteria**



Information criterion (IC) approaches (sometimes called 'Information Theoretic'; IT approaches) can compare all models at once, avoiding the need to cary out multiple pairwise comparisons, and do not require nested models.

In practice, all IC methods reduce to finding the model that minimises some 'criterion' that is the sum of a term based on the likelihood and a penalty term

 $IC \approx \mathcal{L}(\text{model}) + \text{penalty term}$ 

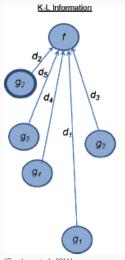
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IC approaches are based on Kullback-Leibler information.

K-L information represents the information lost when model  $g_i$  is used to approximate full reality (f), or the distance between model  $g_i$  and reality.

The goal is then to select the model that minimises K-L information loss (i.e., the model that's closest to reality, f).

The problem is that K-L information loss cannot be computed or estimated.



(Burnham et al., 2011)



In the early 1970s, Hirotogu Akaike made a major breakthrough when he found a formal relationship between K-L information and maximum likelihood (Akaike, 1998).

He focused on the double expectation of the second term of the K-L information and found that, for large sample sizes, this can be estimated simply as  $\ln(\mathcal{L}) - K$ , where K is the total number of estimable parameters in the model.

Akaike multiplied both terms by -2 to get his now famous:

$$AIC = -2\ln(\mathcal{L}) + 2K$$

Note: The term  $-2\ln(\mathcal{L})$  is well known among statisticians as the 'deviance', a goodness-of-fit statistic for a statistical model.



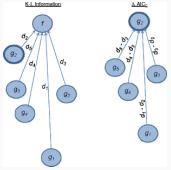
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K-L information represents the distance between model  $g_i$  and reality.

Because we can't estimate 'reality', we instead rely on AIC (or other IC).

We don't know how far the models are from the truth, but their relative positions should be the same, so we can now rank them amongst one another.

Low K-L information means closer to reality, so with AIC, the lower the value the better.



(Burnham et al., 2011)



Akaike's derivations/approximations were based on large sample sizes.

These are not necessarily valid when sample sizes are small.

When the sample sizes are small, there is a good chance that AIC will overfit and select models that have too many parameters.

A small sample size bias correction for AIC was derived that increases the penalty term and is more frequently used in practice. This criterion is denoted as AICc to make it distinct from AIC, and is given by:

AICc = AIC + 
$$\frac{2k^2+2k}{n-k-1}$$

Question: What happens when  $n \to \infty$ ?

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The actual AICc values are not particularly interesting in themselves. It's the differences, or the  $\Delta$ AICc values, that are the key to ranking the models.

When picking among a set of potential models, we select the one with the lowest AICc value of the bunch.

The  $\triangle$ AICc values for any given model are linked to the evidence ratio for the best model as exp[ $-\frac{1}{2}\triangle$ AICc].

For example: A model with a  $\triangle$ AlCc of 2 is  $\sim$ 2.7 times less likely than the best fit model, and a model with a  $\triangle$ AlCc of 50 is 72 billion times less likely

$\triangle AICc$	<b>Evidence</b> Ratio
2	2.7
4	7.4
6	20.1
8	54.6
9	90
10	148.4
11	244
12	403
13	665
14	1,097
15	1,808
20	22,026
50	72 billion

(Burnham et al., 2011)

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The likelihood-ratio can be expressed as a difference between the log-likelihoods:

$$\begin{split} \lambda_{\mathsf{LR}} &= -2(\log \mathcal{L}(\theta_1) - \log \mathcal{L}(\theta_2)) \\ &= -2\log \mathcal{L}(\theta_1) + 2\log \mathcal{L}(\theta_2) \end{split}$$

The  $\triangle$ AIC between a pair of models is

$$\begin{split} \Delta \text{AIC} &= \text{AIC}_{\theta_1} - \text{AIC}_{\theta_2} \\ &= (-2\log\mathcal{L}(\theta_1) + 2K_1) - (-2\log\mathcal{L}(\theta_2) + 2K_2) \\ &= -2\log\mathcal{L}(\theta_1) + 2K_1 + 2\log\mathcal{L}(\theta_2) - 2K_2 \\ &= -2\log\mathcal{L}(\theta_1) + 2\log\mathcal{L}(\theta_2) - 2(K_2 - K_1) \\ \Delta \text{AIC} &= \lambda_{\text{LR}} - 2(K_2 - K_1) \end{split}$$

If  $K_1 = K_2$ , then LRT =  $\Delta$ AIC.



```
library(nlme)
data <- read.csv("Ant Richness.csv")</pre>
FIT <- gls(num_sp ~ latitude + elevation, data = data, method = "ML")
k <- 4 #Intercept, 2 params, and the variance
-2*FIT$logLik + 2*(k)
[1]16.11769
AIC(FIT)
[1]16.11769
AIC(FIT) + (2*k^2 + 2*k)/(nrow(data) - k - 1)
[1] 18,47063
library(MuMIn)
AICc(FIT)
[1] 18,47063
```



```
Let's compare all possible models
FIT <- gls(num_sp ~ latitude + elevation, data = data, method = "HL")
FIT_el <- gls(num_sp ~ elevation, data = data, method = "ML")
FIT_lat <- gls(num_sp ~ latitude, data = data, method = "ML")
INTERCEPT <- gls(num_sp ~ 1, data = data, method = "ML")
AICc(FIT); AICc(FIT_el); AICc(FIT_lat); AICc(INTERCEPT)
[1] 18.47063
[1] 24.67657
[1] 25.04824
[1] 31.07926</pre>
```

This favours the full model. But by how much?

```
      AICc(FIT_e1) - AICc(FIT)
      AICc(INTERCEPT) - AICc(FIT)

      [1]
      6.205939

      1/exp(-(1/2)*6.205939)
      1/exp(-(1/2)*12.60863)

      [1]
      22.26397

      [1]
      546.9268
```

In a paper we would report that the next best model had a  $\triangle$ AlCc of  $\sim$ 6.2, or was  $\sim$ 22 times less likely, and the intercept only model had a  $\triangle$ AlCc of  $\sim$ 12.6 and was >546 less likely than the full model. Biol 520C: Statistical modelling for biological data



# This process is automated by the dredge() function in the ${\tt MuMIn}$ package.

```
FIT <- gls(num_sp ~ latitude + elevation, data = data, method = "ML")
dredge(FIT)
Global model call: gls(model = num_sp ~ latitude + elevation, data = data, method = "ML")
---
Model selection table
(Intrc) elvtn lattd df logLik AICc delta weight
4 11.120 -0.001373 -0.2018 4 -4.059 18.5 0.00 0.922
2 2.489 -0.001613 3 -8.672 24.7 6.21 0.041
3 12.390 -0.2388 3 -8.857 25.0 6.58 0.034
1 2.114 2 -13.224 31.1 12.61 0.002
Models ranked by AICc(x)</pre>
```

Since the advent of AIC, a number of other IC methods have been developed. In essence they all reduce to the combination of some term based on the likelihood and a penalty term based on model complexity.

 $IC \approx \mathcal{L}(\text{model}) + \text{penalty term}$ 

The Bayesian information criterion (BIC) BIC was developed by Gideon Schwarz as an alternative to AIC with a larger penalty term:

$$BIC = -2\ln(\mathcal{L}) + k\ln(n)$$

The Quasi-AIC (QAIC) is an alternative that corrects for overdispersion:

$$QAIC = \frac{-2\ln(\mathcal{L})}{\hat{c}} + 2K$$

When  $\hat{c} = 1$  there is no overdispersion and QAIC = AIC.

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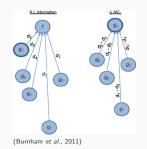
## **IC Considerations**

Because we can't estimate 'reality', IC values only provide relative information (i.e., one model is always going to win out over other models).

Just because a model is the best fit out of a pool of candidates doesn't mean it's any good.

After identifying a candidate model it's important to check all of the assumptions, and test the performance to make sure it's function reasonably well.

For example, comparing with the intercept only model tells us how much of an improvement our model is over simply looking at the mean.







IC are based on a model's likelihood. The likelihood is the probability of some model given data  $\{x\}$ 

#### $\mathcal{L}(\theta|x)$

Models don't need to be nested, but if there are any differences in the datasets, the likelihoods, and therefore the IC, are not comparable (this can be a real problem in practice depending on if your data have NA values, and how they get handled in R).



Sometimes IC based model selection leads to a clear 'winner', other times the differences between the top models are miniscule e.g.  $\Delta$ AlCc of 0.1 (evidence ratio of  $\sim$  1)

What would you do in this situation?

This is where most of the controversy in model selection comes from, and we'll cover some options for handling that situation next lecture.

#### References

- Akaike, H. (1998). Information theory and an extension of the maximum likelihood principle. In: *Selected papers of hirotugu akaike*. Springer, pp. 199–213.
- Burnham, K.P., Anderson, D.R. & Huyvaert, K.P. (2011). AIC model selection and multimodel inference in behavioral ecology: some background, observations, and comparisons. *Behavioral Ecology and Sociobiology*, 65, 23–35.
- Hachoł, J., Hämmerling, M. & Bondar-Nowakowska, E. (2017). Applying the analytical hierarchy process (ahp) into the effects assessment of river training works. *Journal of Water and Land Development*, 35, 63 – 72.
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